TABLE 1 — *Collection localities and museum catalogue number of ants used in this study (TTUno. = ant catalogue number, The Museum, Texas Tech University).*

U.S.A.	Locality	TTUno.
Arizona:	Ruby Road, Sycamore Canyon, Santa Cruz County	6720
Arizona:	Miller Canyon, Huachuca Mountains, Cochise County	6704, 6797
Arizona:	Barfoot Park, Chiracahua Mountains, Cochise County	6942
California:	8 km E. Glenville, Kern County	6783, 6785
Texas:	1.6 km N. Tulsita, Bee County	6992
Texas:	Sam Houston National Forest, Walker County	6994
Texas:	17 km S. Alice, Jim Wells County	6987
Texas:	21 km S. Alice, Jim Wells County	6989
Texas:	37 km S. Tilden, McMullen County	6981, 6991
Texas:	Lubbock, Lubbock County	6982, 6990
Texas:	Post, Garza County	6976

The method of IMAI *et al.* (1977) was followed with minor modifications. Pharate pupae were selected when available and injected with a 0.01% aqueous colchicine solution 18 to 24 hours before dissection following COKENDOLPHER and BROWN (1985). Larvae were occasionally held at room temperature until the pharate pupal stage was reached. After injection with colchicine, the ants were incubated in toweling saturated with 0.01% aqueous colchicine for ca. 24 hours. The brains of the pupae were then removed and placed in a depression slide filled with a hypotonic solution of 1% aqueous sodium citrate for 20 minutes at room temperature. A single brain was then transferred with a Pasteur pipet to a microscope slide and the excess hypotonic solution was drained off. Several drops of fixative 1 (3: 1 glacial acetic acid: absolute ethyl alcohol) were applied and drained away. Two additional drops of fixative were then added and after 10 seconds the brain was minced using extremely fine teasing needles. Two drops of fixative 2 (1: 1 glacial acetic acid: absolute ethyl alcohol) were added and after 30 seconds the fluid was drained away. Several drops of fixative 3 (glacial acetic acid) were applied and after 10 seconds the slide was drained again and set aside to dry overnight. The next day the cells were stained for 10 minutes with 1: 20 Giemsa stain: buffer (COKENDOLPHER and BROWN 1985). The taxa studied and the number of replications of this procedure are presented in Table 2.

The methods of IMAI, BARONI URBANI *et al.* (1984) were followed in the description of chromosome morphologies. In that system, chromosomes are grouped into categories: Ā (acrocentric in a broad sense, which includes traditional T and A) and M̄ (metacentric in broad sense, which includes traditional M; SM and ST).

RESULTS AND DISCUSSION

The results of the karyological survey are summarized in Table 2. The detailed descriptions of the karyotypes are as follows: