

- 2b. Body any one of diverse shapes but never as above; temporal sulci absent; antennae usually on upper half of cranium . . . . . Formicidae
- 3a. Body straight; diameter greatest at AI and AII; gradually attenuated toward posterior end and more rapidly toward anterior end; each in a separate cell of a paper comb; usually hanging head down . . . . . Vespidae
- 3b. Body stout, not straight; not in ~~separate~~ <sup>paper</sup> cells . . . . . 4
- 4a. Thorax and AI bent ventrally; abdomen straight and of nearly uniform diameter; opening of sericteries without lips; nests in soil or rotten wood; cells lined with wax . . . . . Halictidae
- 4b. Body crescentric; opening of sericteries with conspicuous lips; cells constructed of wax or of a mixture of wax and pollen . . . . . Apidae

#### HISTORY AND METHODS (p. 1)

We now prefer 70% alcohol. If tax-free alcohol is not available, rubbing alcohol (ethyl but *not* isopropyl) may be used.

Drawing (p. 1).—Line drawings are not difficult to make: see our 1960a. Inking, however, does require more skill, which can be acquired with practice. We now use polyester drafting film, India ink for film and Rapidograph drawing pens for film (sizes 000, 0 and 1); the Hunt mapping pen is still good for details such as hairs on the body. The great advantage of film is that errors can be erased or carefully scraped off with a knife. French curves are useful but—as usual—must be carefully fitted to the desired curve.

The Scanning Electron Microscope (p. 1).—Photographs made by a scanning electron microscope (SEM) are now popular, although their use sometimes appears to be superfluous. In this regard, we cite the article by Clark and Glavog (1976: 1361). SEMs are especially useful when it is desirable to magnify higher than can be done with a light microscope, e.g. surface features of the integument. We find that any whole larva or any detail that can be seen under a light microscope can be shown more clearly with a photomicrograph or a line drawing. An example of the superiority of line drawings over SEMs is to be found in Kempf (1975).

#### LITERATURE (p. 1)

As the next-to-last sentence in the first paragraph insert “They total 1298.”  
In the last line change 577 to 632.

#### MATERIAL STUDIED (p. 1)

We have studied the larvae of 777 species in 202 genera representing 51 of the 61 tribes and all 10 of the living subfamilies. The taxa are given in Appendix B. A summary by subfamilies of the number of genera and species (in parentheses) follows: Dorylinae 6 (23); Leptanillinae 2 (3); Cerapachyinae 5 (11); Myrmeciinae 2 (31); Ponerinae 41 (169); Pseudomyrmecinae 4 (34); Myrmicinae 94 (299); Aneuretinae 1 (1); Dolichoderinae 14 (54); Formicinae 33 (154).

#### GEOGRAPHICAL DISTRIBUTION (p. 2)

Additions only.—AFRICA—Angola 2, Cameroon 2, Ghana 3, Ivory Coast 5, Kenya 3, Madagascar 5, Morocco 1, South Africa 2, Tunisia 1, Zambia 1.

ASIA—India 4, Japan 3, Malaya 1, Singapore 1.